STA305/1004-Class 17

Nov. 21, 2019

Today's Class

ANOVA

- Multiple comparisons
- Sample size for ANOVA

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- 3. $\mu_2 \neq \mu_3$

all pairwise comparisons.

► Suppose that k = 3 separate (independent) hypothesis tests at level α tests are conducted: $H_{0_k} : \mu_i = \mu_j \vee s$. $H_{1_k} : \mu_i \neq \mu_j$.

- Suppose that k = 3 separate (independent) hypothesis tests at level α tests are conducted: H_{0k} : μ_i = μ_j vs. H_{1k} : μ_i ≠ μ_j.
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$$A_{K} = \{ veject H_{0K} \}$$

$$P (reject at (east one H_{0K}) = P \begin{pmatrix} 3 \\ 0 \\ 0 \\ 0 \end{pmatrix}$$

$$= A^{C} \cap B^{C} \qquad = I - P \begin{pmatrix} (3 \\ 0 \\ 0 \\ 0 \\ 0 \end{pmatrix}$$

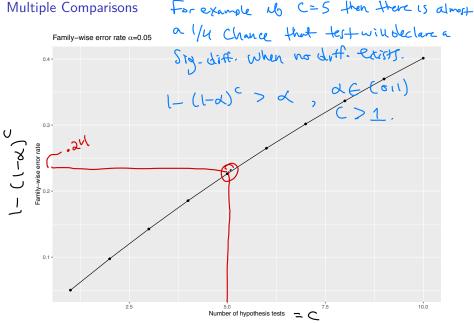
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$$H_{0_k}$$
) =
1 - P (do not reject H_{0_1} and do not reject H_{0_2} and do not reject H_{0_3})
L - P (A, c) P(A, c) P(A_3 c) Au = {Veyect H_{0_k}]
assuming that A₂, A₂, A₃
are independent.
= L - (L-a)(L-a)(L-a) = L - (L-a)³

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- Since the hypotheses are independent: $1-P(\text{do not reject } H_{0_1})P(\text{do not reject } H_{0_2})P(\text{do not reject } H_{0_3}) = 1-(1-\alpha)^3$
- If $\alpha = 0.05$ then the probability that at least one H_0 will be falsely rejected is $1 (1 .05)^3 = 0.14$, which is almost three times the type I error rate.



$$H_0: \mu_1 = \mu_2 = \cdots = \mu_k \, \mathsf{vs.} \, H_1: \mu_i \neq \mu_j.$$

If c independent hypotheses are conducted then the probability

P
$$\left(\mathsf{reject} \,\, \mathsf{at} \,\, \mathsf{least} \,\, \mathsf{one} \,\, {\mathcal{H}_{0}}_k
ight) = 1 - (1 - lpha)^c$$

is called the family-wise error rate.

The pairwise error rate is $P(\text{reject } H_{0_k}) = \alpha$ for any c.

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- If treatment means are significantly different from the ANOVA F test then researchers usually want to explore which means are different.
- Is it appropriate to test for differences looking at all pairwise comparisons?
- Testing all possible pairs increases the type I error rate.
- This means that there is a higher probability, beyond the pre-stated type I error rate (e.g. 0.05), that that a significant difference is detected when the truth is that no difference exists.

Example



Neural correlates of interspecies perspective taking in the post-mortem Atlantic Salmon: An argument for multiple comparisons correction

Craig M. Bennett¹, Abigail A. Baird², Michael B. Miller¹, and George L. Wolford³ ¹Psychology Department, University of Californii Santa Barbara, Saria Barbara, CA¹² Department of Psychologial 8 min Selores, Datamosh Collega, Nanoer, NH

INTRODUCTION

With the extreme dimensionality of functional neuroimaging data comes extreme risk for faile positives. Across the 130,000 voxels in a typical IMRI volume the probability of a faile positive is almost certain. Correction for multiple comparisons should be completed with these datasets, but is often ignored by investigators. To illustrate the magnitude of the problem we carried out a real experiment that demonstrates the datager of not correcting for chance recerverty.

METHODS

Subject. One mature Atlantic Salmon (Salmo salar) participated in the fMRI study. The salmon was approximately 18 inches long, weighed 3.8 lbs, and was not alive at the time of scanning.

Task. The task administered to the solmon involved completing an open-ended metallicing task. The salmen was shown a senise of photographs depicting human individuals in social sinustions with a specified metricean valence. The salmen was asked to determine what errotion the individual in the photo must have been experimently.

Design. Stimuli were presented in a block design with each photo presented for 10 seconds followed by 12 seconds of rest. A total of 15 photos were displayed. Total scan time was 5.5 minutes.

Progreeossing, Image processing was completed using SPM2. Preprocessing steps for the functional imaging data included a 6-parameter rigid-body affine realignment of the fMR1 timeseries, corregistration of the data to a T₁-weighted matornical image, and 8 mm full-width at half-maximum (FWHM) Gaussian smoothing.

<u>Analysis</u>, Vostvivie statistics on the salmon data were calculated through an ordinary last-square solimation of the general linear model (GIAM). Predicters of the henodynamic response, were modeled by a bocase function convolved with a control henodynamic response. A temporal high pass filter of 128 seconds was include to account fee low frequency drift. No autocorrelation correction was applied.

<u>Yanti Section</u>, Two methods were used for the correstions of multiple comparisons in the DNRI result. The first method controlled the overall faile discovery rate (PDR) and was based on a method defined by Bergiumin and Hostberg (1995). The second method controlled the overall finallysisses are rate rate (PWRR) through the use of Gaussian madem field theory. This was done using algorithms originally devised by Friston et al. (1994).

DISCUSSION

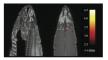
Cas we conclude from this data that the subnox is engapsing in the properior-testing start Certainly sort. When we can determine it that random noise in the IIP functories may yield sparines results if multiples comparisons are accellent options and we will be well-this in all mark TMM analysis packages. We input the relying on standard statistical thresholds (q < 0.001) and som miximum data using (h < 0.01) and the full starking in comparisons. We listificate same that the box out majority of MRR studies food comparations. We inform statistical thresholds (q < 0.001) and the comparation of the statistical thresholds (q < 0.001) and the comparation of the statistical thresholds (q < 0.001) and the comparation of the statistical.

REFERENCES

Benjamini Y and Hochberg Y (1995). Controlling the false discovery rate: a practical and powerful approach to multiple tosting. Journal of the Royal Statistical Society: Series 8, 57:289-303.

Fristen KJ, Worsley KJ, Frackowisk RSJ, Mazziotta JC, and Evans AC. (1994). Assessing the significance of focal activations using their spatial extent. *Huware Brain Mapping*, 1:214-228.

GLM RESULTS

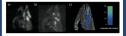


A *t*-contrast was used to test for regions with significant BOLD signal change during the photo condition compared to rest. The parameters for this comparison were n(131) > 3.15, p(uncorrected) < 0.001, 3 voxel extent threshold.

Several active vocetis were discovered in a cluster located within the salmore born active (Figure 1, see above). The size of this cluster was 81 cm² with a cluster-level significance of p = 0.001. Due to the coarse resolution of the co-hop-tanar image acquisition number levelatively small size of the salmon brain further discrimination between brain regions could not be completed. Out of a search volume of 5040 vocuses is not all of two outsile were significant.

Identical t-contrasts controlling the false discovery rate (FDR) and familywise error rate (FWER) were completed. These contrasts indicated no active vectels, even at relaxed statistical thresholds (n = 0.25).

VOXELWISE VARIABILITY



To examine the spatial configuration of false positives we completed a variability analysis of the fMRI timeseries. On a voxel-by-voxel basis we calculated the standard deviation of signal values across all 140 volumes.

We observed clustering of highly variable vocels into groups near areas of high voxel signal intensity. Figure 2a shows the mean EPI image for all 140 image volumes. Figure 2b shows the standard deviation values of each voxel. Figure 2c shows thresholded standard deviation values overlaid ento a highresolution T, vesighted image.

To investigate this effect in groater detail we conducted a Pearson correlation to examine the relationship between the signal in a vocel and its variability. There was a significant positive correlation between the mean positive correlation between the mean statistical or the significant positive vocel value and its variability over time (r = 0.54, p < 0.001). A scatterplied of mean vocel signal intensity against voxel standard deviation is presented to the right.



The Bonferroni Method

To test for the difference between the *i*th and *j*th treatments, it is common to use the two-sample t test. The two-sample t statistic is

$$t_{ij} = rac{y_{ar{j}\cdot} - y_{ar{i}\cdot}}{\hat{\sigma}\sqrt{1/n_j + 1/n_i}},$$

where y_{j} . is the average of the n_i observations for treatment j and $\hat{\sigma}$ is $\sqrt{MS_E}$ from the ANOVA table.

Treatments *i* and *j* are declared significantly different at level α if

$$|t_{ij}| > t_{N-k,\alpha/2},$$
 Critical value.

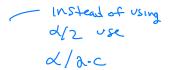
where $t_{N-k,\alpha/2}$ is the upper $\alpha/2$ percentile of a t_{N-k} .

The total number of pairs of treatment means that can be tested is

$$c = {\binom{k}{2}} = \frac{k(k-1)}{2}.$$

The Bonferroni method for testing $H_0: \mu_i = \mu_j$ vs. $H_0: \mu_i \neq \mu_j$ rejects H_0 at level α if

$$|t_{ij}| > t_{N-k,\alpha/2c},$$

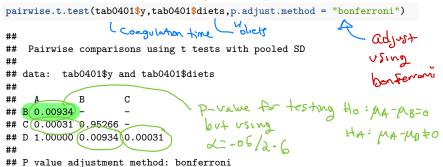


where *c* denotes the number of pairs being tested.

The Bonferroni Method

In R the function pairwise.t.test() can be used to compute Bonferroni adjusted p-values.

This is illustrated below for the blood coagualtion study.

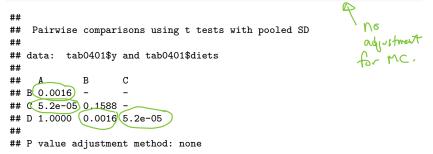


There are significant differences at the 5% level between diets A and B, A and C, B and D, and C and D using the Bonferroni method.

The Bonferroni Method

For comparison the unadjusted p-values are also calculated.

pairwise.t.test(tab0401\$y,tab0401\$diets,p.adjust.method = "none")



The significant differences are the same using the unadjusted p-values but the p-values are larger then the p-values adjusted using the Bonferroni method.

A 100 $(1 - \alpha)$ % simultaneous confidence interval for c pairs $\mu_i - \mu_i$ is

$$\bar{y_{j\cdot}} - \bar{y_{i\cdot}} \pm t_{N-k} \alpha/2c} \hat{\sigma} \sqrt{1/n_j + 1/n_i}.$$

After identifying which pairs are different, the confidence interval quantifies the range of plausible values for the differences.

The Bonferroni Method - coagulation study

The treatment means can be obtained from the table below.

	А	В	С	D
	60	65	71	62
	63	66	66	60
	59	67	68	61
	63	63	68	64
	62	64	67	63
	59	71	68	56
Treatment Average	61	66	68	61
Grand Average	64	64	64	64
Difference	-3	2	4	-3

The Bonferroni Method - coagulation study

```
\hat{\sigma} = \sqrt{MS_E} can be obtained from the ANOVA table.
anova(lm(y~diets,data=tab0401))
```

```
## Analysis of Variance Table
##
## Response: y
##
             Df Sum Sq Mean Sq F value
                                            Pr(>F)
              3
                    228
                           76.0 13.571 4.658e-05 ***
## diets
## Residuals 20
                    112
                            5.6
## ---
                   0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## Signif. codes:
The upper .05/(2 \cdot 6) = 0.004 percentile of the t_{24-4} can be obtained with the t
quantile function in R qt().
                                          24-4
qt(p = 1-0.004, df = 20)
                                                                1-004
                                    tro
## [1] 2.945349
                                                                     004
                                                                 2011
```

The Bonferroni Method - coagulation study

Plugging in these values to the confidence interval formula we can obtain a Bonferroni adjusted 95% confidence interval for $\mu_B - \mu_A$:

$$66-61\pm 2.95\sqrt{5.6}\sqrt{1/6+1/6}$$

The lower and upper limits can be calculated in R. 66-61 - qt(p = 1-0.004, df = 20)*sqrt(5.6)*sqrt(1/6+1/6) # lower limit

[1] 0.9758869
66-61 + qt(p = 1-0.004,df = 20)*sqrt(5.6)*sqrt(1/6+1/6) # upper limit

[1] 9.024113

The 95% confidence interval for $\mu_B - \mu_A$ is (0.98, 9.02).

Adjusted for Mr using Bonfernni

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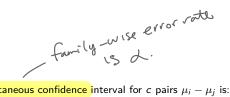
$$t_{ij} \text{ is the observed value of the two-sample t-statistic}$$

$$q_{k,N-k,\alpha} \text{ is the upper } \alpha \text{ percentile of the Studentized range distribution with parameters } k \text{ and } N-k \text{ degrees of freedom.}$$

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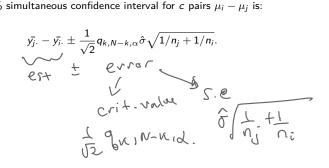
- t_{ij} is the observed value of the two-sample t-statistic
- $q_{k,N-k,\alpha}$ is the upper α percentile of the Studentized range distribution with parameters k and N-k degrees of freedom.
- The CDF and inverse CDF of the Studentized Range Distribution is available in R via the functions ptukey() and qtukey() respectively.



• A $100(1-\alpha)$ % simultaneous confidence interval for c pairs $\mu_i - \mu_j$ is:

►

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$$y_{\overline{j}\cdot} - y_{\overline{i}\cdot} \pm \frac{1}{\sqrt{2}} q_{k,N-k,\alpha} \hat{\sigma} \sqrt{1/n_j + 1/n_i}.$$

The Bonferroni method is more conservative than Tukey's method. In other words, the simutaneous confidence intervals based on the Tukey method are shorter.

► In the coagualtion study N = 24, k = 4 so the 5% critical value of the Studentized range distribution is obtained using the the inverse CDF function qtukey() for this distribution.

Tivey.

• The argument lower.tail=FALSE is used so we obtain the upper percentile of the distribution (i.e., the value of x such that P(X > x) = 0.05).

qtukey(p = .05,nmeans = 4,df = 20,lower.tail = FALSE)

[1] 3.958293

Let's obtain the Tukey p-value and confidence interval for $\mu_B-\mu_A.$ The observed value of the test statistic is:

$$q^{obs} = \sqrt{2}|t_{AB}|, \qquad - \text{The Turkey procedure}$$

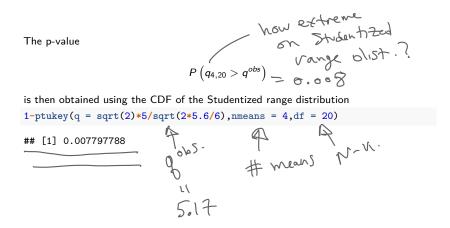
$$t_{AB} = \frac{y_{\overline{A}} - y_{\overline{B}}}{\hat{\sigma}\sqrt{1/n_A + 1/n_B}}. \qquad \text{Varge of ist. to}$$

$$respective of the statement of the statem$$

where

(sqrt(2)*(6

[1] 5.17



The 95% limits of the Tukey confidence interval for $\mu_B - \mu_A$ is tuk.crit <- qtukey(p=.05,nmeans=4,df=20,lower.tail=FALSE) #lower limit round(5-(1/sqrt(2))*tuk.crit*sqrt(5.6)*sqrt(1/6+1/6),2) ## [1] 1.18 $e_{S+} - e_{rror}$ #upper limit round(5+(1/sqrt(2))*tuk.crit*sqrt(5.6)*sqrt(1/6+1/6),2) ## [1] 8.82 $e_{S+} + e_{rror}$

The width of the Tukey confidence interval for $\mu_B - \mu_A$ is round((1/sqrt(2))*tuk.crit*sqrt(5.6)*sqrt(1/6+1/6),2)

[1] 3.82
The width of Bonferroni µ_B - µ_A is
round(qt(p = 1-0.004,df = 20)*sqrt(5.6)*sqrt(1/6+1/6),2)
[1] 4.02

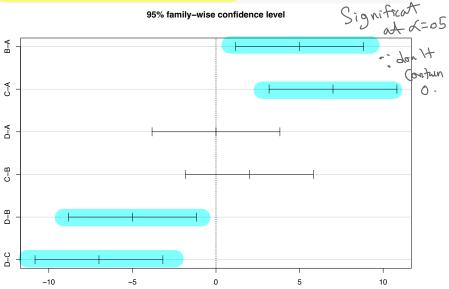
- This shows that the Tukey confidence interval is shorter than Bonferroni confidence intervals.
- The command TukeyHSD() can be used to obtain all the Tukey confidence intervals and p-values for an ANOVA.

TukeyHSD(aov(y~diets,data=tab0401))

round(TukeyHSD(aov(y~diets,data=tab0401))\$diets,2)

##		diff	lwr	upr	p adj			
##	B-A	5	1.18	8.82	0.01	æ		
##	C-A	7	3.18	10.82	0.00	*		
##	D-A	0	-3.82	3.82	1.00			
##	C-B	2	-1.82	5.82	0.48			
##	D-B	-5	-8.82	-1.18	0.01	¥		
##	D-C	-7	-10.82	-3.18	0.00	*		
				4				-
				\subseteq	. 610e	s not	Contain	O





Differences in mean levels of diets